

УДК 577.21

ИНАКТИВАЦИЯ Ras1 В ДЕЛЯЩИХСЯ ДРОЖЖАХ УСИЛИВАЕТ ОТВЕТ НА ОКИСЛИТЕЛЬНЫЙ СТРЕСС, ИНДУЦИРУЕМЫЙ *тrem*-БУТИЛГИДРОПЕРОКСИДОМ (tBHP)¹

© 2023 г. N. Masood^a, S. Anjum^b, S. Ahmed^b, *

^aBiochemistry and Structural Biology Division, CSIR- Central Drug Research Institute, Sector 10, Jankipuram Extension, Sitapur Road, Lucknow, 226031 India

^bAcademy of Scientific and Innovative Research (AcSIR), Ghaziabad, 201002 India

*e-mail: shakil_ahmed@cdri.res.in

Поступила в редакцию 22.07.2022 г.

После доработки 29.11.2022 г.

Принята к публикации 24.12.2022 г.

Малые GTPазы Ras функционируют как молекулярные переключатели, регулирующие клеточный гомеостаз. Ras-зависимые сигнальные пути регулируют такие важные процессы, как прохождение клеточного цикла, апоптоз, миграция и старение клеток. Нарушение сигнального пути Ras связано с несколькими патологическими состояниями. Установлено, что белки Ras могут участвовать в регуляции окислительно-восстановительных сигнальных путей, включая влияние на уровень активных форм кислорода, создающих условия для канцерогенеза. Предполагается, что активные формы кислорода и разобщение митохондриальных функций являются главными факторами, воздействующими на физиологические процессы в клетках и вовлеченными в разные патологии. В настоящей работе изучена роль Ras1, *тrem*-бутилгидропероксида (tBHP) и антимицина А в ответе клеток *Schizosaccharomyces pombe* на окислительный стресс. Обнаружено снижение выживаемости, более высокий уровень активных форм кислорода и нарушение функций митохондрий в клетках *ras1Δ* и в клетках дикого типа, обработанных tBHP, а также ингибитором дыхательной цепи антимицином А. Более того, эти эффекты сильнее выражены в обработанных антимицином или tBHP клетках *ras1Δ*. Показано также, что Ras1 регулирует экспрессию и активность таких антиоксидантных ферментов, как глутатионпероксидаза (GSH-Px), глутатион-S-трансфераза (GST) и каталаза. Эти результаты свидетельствуют о потенциальной роли Ras1 *S. pombe* в смягчении ответа на окислительный стресс.

Ключевые слова: *Schizosaccharomyces pombe*, Ras1, активные формы кислорода, ROS, *тrem*-бутилгидропероксид tBHP, окислительный стресс

DOI: 10.31857/S0026898423040158, EDN: QLULLQ

СПИСОК ЛИТЕРАТУРЫ

1. Sbodio J.I., Snyder S.H., Paul B.D. (2019) Redox mechanism in neurodegeneration: from disease outcome to therapeutic opportunities. *Antioxid. Redox Signal.* **30**, 1450–1499.
2. Simanshu D.K., Nissley D.V., McCormick F. (2017) RAS proteins and their regulators in human disease. *Cell.* **170**(1), 17–33.
3. Vojtek A.B., Der C.J. (1998) Increasing complexity of the Ras signaling pathway. *J. Biol. Chem.* **273**, 19925–19928.
4. Klandorf H., Dyke Van K. (2012) Oxidative and nitrosative stresses: their role in health and disease in man and birds. In: *Oxidative Stress – Molecular Mechanisms and Biological Effects*. Eds Lushchak V., Semchyshyn H.M. IntechOpen, 47–60. <https://doi.org/10.5772/33879>
5. Carew J.S., Zhou Y., Huang P. (2006) Oxidative stress, cell proliferation, and apoptosis. In: *Oxidative Stress, Disease and Cancer*. Ed. Singh K.K. London: Imperial College Press, 309–331. https://doi.org/10.1142/9781860948046_0009
6. Marozkina N.V., Gaston B. (2012) S-Nitrosylation signaling regulates cellular protein interactions. *Biochim. Biophys. Acta.* **1820**, 722–729.
7. Malumbres M., Barbacid M. (2003) RAS oncogenes: the first 30 years. *Nat. Rev. Cancer.* **3**, 459–465.
8. Garcia P., Tajadura V., Garcia I., Sanchez Y. (2006) Role of Rho GTPases and Rho GEFs in the regulation of cell shape and integrity in fission yeast. *Yeast.* **23**, 1031–1043.

¹ Статья представлена авторами на английском языке.

9. Young E., Zheng Z.Y., Wilkins A.D., Jeong H.T., Li M., Lichtarge O., Chang E.C. (2014) Regulation of Ras localization and cell transformation by evolutionarily conserved palmitoyltransferases. *Mol. Cell. Biol.* **34**, 374–385.
10. Kim H.J., Jung H.Y., Lim C.J. (2008) The pap1+ gene of fission yeast is transcriptionally regulated by nitrosative and nutritional stress. *FEMS Microbiol. Lett.* **280**, 176–181.
11. Bond M., Croft W., Tyson R., Bretschneider T., Davey J., Ladds G. (2013) Quantitative analysis of human ras localization and function in the fission yeast *Schizosaccharomyces pombe*. *Yeast*. **30**, 145–156.
12. Weston C., Bond M., Croft W., Ladds G. (2013) The coordination of cell growth during fission yeast mating requires Ras1-GTP hydrolysis. *PLoS One*. **8**(10), e77487
13. Sánchez N.S., Königsberg M. (2006) Using yeast to easily determine mitochondrial functionality with 1-(4,5-dimethylthiazol-2-yl)-3,5-diphenyltetrazolium bromide (MTT) assay. *Biochem. Mol. Biol. Educ.* **34**(3), 209–212.
14. Warholm M., Guthenberg C., von Bahr C., Mannervik B. (1985) Glutathione transferases from human liver. *Methods Enzymol.* **113**, 499–504.
15. Wendel A. (1981) Glutathione peroxidase. *Methods Enzymol.* **77**, 325–333.
16. Roggenkamp R., Sahm H., Wagner F. (1974) Microbial assimilation of methanol induction and function of catalase in *Candida boidinii*. *FEBS Lett.* **41**(2), 283–286.
17. Vlamis-Gardikas A., Åslund F., Spyrou G., Bergman T., Holmgren A. (1997) Cloning, overexpression, and characterization of glutaredoxin 2, an atypical glutaredoxin from *Escherichia coli*. *J. Biol. Chem.* **272**(17), 11236–11243.
18. Sonkar A., Yadav S., Ahmed S. (2016) Cleavage and polyadenylation factor, Rna14 is an essential protein required for the maintenance of genomic integrity in fission yeast *Schizosaccharomyces pombe*. *Biochim. Biophys. Acta* **1863** (2), 189–197.
19. Amoroso S., D'Alessio A., Sirabella R., Di Renzo G., Annunziato L. (2002) Ca(2+) independent caspase-3 but not Ca2+-dependent caspase-2 activation induced by oxidative stress leads to SH-SY5Y human neuroblastoma cell apoptosis. *J. Neurosci. Res.* **68**, 454–462.
20. Kanupriya A., Prasad D., Sai Ram M., Sawhney R.C., Ilavazhagan G., Banerjee P.K. (2007) Mechanism of tert-butylhydroperoxide induced cytotoxicity in U-937 macrophages by alteration of mitochondrial function and generation of ROS. *Toxicol. In Vitro*. **21**(5), 846–854.
21. Lv H., Zhen C., Liu J., Yang P., Hu L., Shang P. (2019) Unraveling the potential role of glutathione in multiple forms of cell death in cancer therapy. *Oxid. Med. Cell. Longev.* **2019**, 3150145.
22. Auten R.L., Davis J.M. (2009) Oxygen toxicity and reactive oxygen species: the devil is in the details. *Pediatr. Res.* **66**(2), 121–127.
23. Guo C., Sun L., Chen X., Zhang D. (2013) Oxidative stress, mitochondrial damage and neurodegenerative diseases. *Neural. Regen. Res.* **8**(21), 2003.
24. Bhatti J.S., Bhatti G.K., Reddy P.H. (2017) Mitochondrial dysfunction and oxidative stress in metabolic disorders — a step towards mitochondria based therapeutic strategies. *Biochim. Biophys. Acta. Mol. Basis Dis.* **1863**(5), 1066–1077.
25. Veal E.A., Toone W.M., Jones N., Morgan B.A. (2002) Distinct roles for glutathione S-transferases in the oxidative stress response in *Schizosaccharomyces pombe*. *J. Biol. Chem.* **277**, 35523–35531.
26. Aniya Y., Daido A. (1994) Activation of microsomal glutathione S-transferase in tert-butyl hydroperoxide-induced oxidative stress of isolated rat liver. *Jpn. J. Pharmacol.* **66**(1), 123–130.
27. Takebe G., Yarimizu J., Saito Y., Hayashi T., Nakamura H., Yodoi J., Nagasawa S., Takahashi K. (2002) A comparative study on the hydroperoxide and thiol specificity of the glutathione peroxidase family and selenoprotein P. *J. Biol. Chem.* **277**(43), 41254–41258.
28. Inoue Y., Matsuda T., Sugiyama K.I., Izawa S., Kimura A. (1999) Genetic analysis of glutathione peroxidase in oxidative stress response of *Saccharomyces cerevisiae*. *J. Biol. Chem.* **274**(38), 27002–27009.
29. Sandström B.E., Marklund S.L. (1990) Effects of variation in glutathione peroxidase activity on DNA damage and cell survival in human cells exposed to hydrogen peroxide and *t*-butyl hydroperoxide. *Biochem. J.* **271**(1), 17–23.
30. Jamieson D.J. (1998) Oxidative stress responses of the yeast *Saccharomyces cerevisiae*. *Yeast*. **14**(16), 1511–1527.
31. Zhang J., Wang X., Vikash V., Ye Q., Wu D., Liu Y., Dong W. (2016) ROS and ROS-mediated cellular signaling. *Oxid. Med. Cell. Longev.* **2016**, 4350965. <https://doi.org/10.1155/2016/4350965>
32. Cox A. D., Der C. J. (2003) The dark side of Ras: regulation of apoptosis. *Oncogene*. **22**, 8999–9006.
33. Shaulian E., Karin M. (2001) AP-1 in cell proliferation and survival. *Oncogene*. **20**, 2390–2400.
34. Weinberg F., Hamanaka R., Wheaton W.W., Weinberg S., Joseph J., Lopez M., Kalyanaraman B., Mutlu G.M., Budinger G.S., Chandel N.S. (2010) Mitochondrial metabolism and ROS generation are essential for Kras-mediated tumorigenicity. *Proc. Natl. Acad. Sci. USA*. **107**(19), 8788–8793.
35. Toone W.M., Kuge S., Samuels M., Morgan B.A., Toda T., Jones N. (1998) Regulation of the fission yeast transcription factor Pap1 by oxidative stress: requirement for the nuclear export factor Crm1 (Exportin) and the stress-activated MAP kinase Sty1/Spc1. *Genes Dev.* **12**(10), 1453–1463.
36. Lim J.K., Delaidelli A., Minaker S.W., Zhang H.F., Colovic M., Yang H., Negri G.L., von Karstedt S., Lockwood W.W., Schaffer P., Leprivier G. (2019) Cystine/glutamate antiporter xCT (SLC7A11) facilitates oncogenic RAS transformation by preserving intracellular redox balance. *Proc. Natl. Acad. Sci. USA*. **116**, 9433–9442.
37. Padanad M.S., Konstantinidou G., Venkateswaran N., Melegari M., Rindhe S., Mitsche M., Yang C., Batten K., Huffman K.E., Liu J., Tang X. (2016) Fatty acid oxidation mediated by Acyl-CoA synthetase long chain 3 is required for mutant KRAS lung tumorigenesis. *Cell Rep.* **16**, 1614–1628.
38. Carracedo A., Cantley L.C., Pandolfi P.P. (2013) Cancer metabolism: fatty acid oxidation in the limelight. *Nat. Rev. Cancer*. **13**, 227–232.

Inactivation of Ras1 in Fission Yeast Aggravates the Oxidative Stress Response Induced by Tert Butyl Hydroperoxide (tBHP)

N. Masood¹, S. Anjum², and S. Ahmed^{2, *}

¹Biochemistry and Structural Biology Division, CSIR- Central Drug Research Institute, Sector 10, Jankipuram Extension, Sitapur Road, Lucknow, 226031 India

²Academy of Scientific and Innovative Research (AcSIR), Ghaziabad, 201002 India

*e-mail: shakil_ahmed@cdri.res.in

Ras proteins are small GTPases and function as molecular switches to regulate cellular homeostasis. Ras-dependent signalling pathways regulate several essential processes such as cell cycle progression, growth, migration, apoptosis, and senescence. The dysregulation of Ras signaling pathway has been linked to several pathological outcomes. A potential role of RAS in regulating the redox signalling pathway has been established that includes the manipulation of ROS levels to provide a redox milieu that might be conducive to carcinogenesis. Reactive oxygen species (ROS) and mitochondrial impairment have been proposed as major factors affecting the physiology of cells and implicated in several pathologies. The present study was conducted to evaluate the role of Ras1, tert Butyl hydroperoxide (tBHP), and antimycin A in oxidative stress response in *Schizosaccharomyces pombe* cells. We observed decreased cell survival, higher levels of ROS, and mitochondrial dysfunctionality in *ras1Δ* cells and tBHP as well as respiratory inhibitor, antimycin A treated wild type cells. Furthermore, these defects were more profound in *ras1Δ* cells treated with tBHP or antimycin A. Additionally, Ras1 also has been shown to regulate the expression and activity of several antioxidant enzymes like glutathione peroxidase (GSH-Px), glutathione-S-transferase (GST), and catalase. Together, these results suggest the potential role of *S. pombe* Ras1 in mitigating oxidative stress response.

Keywords: *Schizosaccharomyces pombe*, Ras1, ROS, tBHP, oxidative stress