

СТРУКТУРНО-ФУНКЦИОНАЛЬНЫЙ АНАЛИЗ БИОПОЛИМЕРОВ И ИХ КОМПЛЕКСОВ

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РАЗДЕЛЕНИЕ ФАЗ В ПРЕПАРАТЕ ОЧИЩЕННОГО БЕЛКА ЧЕЛОВЕКА LSM4¹

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Жидкофазное разделение белков происходит в ряде биологических процессов, таких как регуляция транскрипции, процессинг и созревание РНК. Sm-подобный белок 4 (LSM4) участвует во множестве процессов, включая сплайсинг пре-мРНК и сборку Р-телец. Перед проведением исследований по участию LSM4 в разделении двух жидких фаз в ходе процессинга или созревания РНК необходимо сначала обнаружить эту способность белка LSM4 в системе *in vitro*. Плазмиду, экспрессирующую белок mCherry-LSM4, получали на основе вектора pET30a и использовали для выделения белка mCherry-LSM4 из прокариотических клеток (*Escherichia coli* штамм BL21). Белок mCherry-LSM4 очищали с использованием аффинной хроматографии на Ni-NTA-агарозе, а затем жидкостной хроматографии быстрого разрешения (FPLC). Широкопольную флуоресцентную микроскопию Delta-Vision использовали для наблюдения за динамическим разделением фаз жидкость–жидкость в препаратах белка LSM4 *in vitro*. В ходе анализа структуры белка LSM4 с помощью базы данных Predictor of Natural Disordered Regions установлено, что его С-конец содержит домен низкой сложности. Из *E. coli* получен очищенный препарат полноразмерного белка LSM4 человека. Показано, что LSM4 человека обеспечивает зависимое от концентрации разделение фаз жидкость–жидкость *in vitro* в буфере с краудинг-агентами. Соли в высокой концентрации и 1,6-гександиол блокируют LSM4-индуцированное разделение двух жидких фаз. Кроме того, *in vitro* наблюдается слияние капель белка LSM4. Полученные результаты свидетельствуют о том, что полноразмерный белок LSM4 человека обладает способностью образовывать жидкие включения и вызывать разделение фаз жидкость–жидкость *in vitro*.

Ключевые слова: разделение фаз жидкость–жидкость, Sm-подобный белок 4, жидкостная экспресс-хроматография белков, очистка белков, биофизический процесс

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СПИСОК ЛИТЕРАТУРЫ

1. Banani S.F., Lee H.O., Hyman A.A., Rosen M.K. (2017) Biomolecular condensates: organizers of cellular biochemistry. *Nat. Rev. Mol. Cell. Biol.* **18**(5), 285–298.
2. Mitrea D.M., Cika J.A., Stanley C.B., Nourse A., Onuchic P.L., Banerjee P.R., Phillips A.H., Park C.G., Deniz A.A., Kriwacki R.W. (2018) Self-interaction of NPM1 modulates multiple mechanisms of liquid–liquid phase separation. *Nat. Commun.* **9**(1), 842.
3. McSwiggen D.T., Hansen A.S., Teves S.S., Marie-Nelly H., Hao Y., Heckert A.B., Umemoto K.K., Dugast-Darzacq C., Tjian R., Darzacq X. (2019) Evidence for DNA-mediated nuclear compartmentalization distinct from phase separation. *Elife*. **8**, e47098.

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4. Hyman A.A., Weber C.A., Julicher F. (2014) Liquid-liquid phase separation in biology. *Annu. Rev. Cell. Dev. Biol.* **30**, 39–58.
5. Aguzzi A., Altmeyer M. (2016) Phase separation: linking cellular compartmentalization to disease. *Trends Cell Biol.* **26**(7), 547–558.
6. Bergeron-Sandoval L.P., Safaei N., Michnick S.W. (2016) Mechanisms and consequences of macromolecular phase separation. *Cell.* **165**(5), 1067–1079.
7. Toretzky J.A., Wright P.E. (2014) Assemblages: functional units formed by cellular phase separation. *J. Cell Biol.* **206**(5), 579–588.
8. Lyon A.S., Peeples W.B., Rosen M.K. (2021) A framework for understanding the functions of biomolecular condensates across scales. *Nat. Rev. Mol. Cell Biol.* **22**(3), 215–235.
9. Sanulli S., Trnka M.J., Dharmarajan V., Tibble R.W., Pascal B.D., Burlingame A.L., Griffin P.R., Gross J.D., Narlikar G.J. (2019) HP1 reshapes nucleosome core to promote phase separation of heterochromatin. *Nature.* **575**(7782), 390–394.
10. Zhao Y.G., Zhang H. (2020) Phase separation in membrane biology: the interplay between membrane-bound organelles and membraneless condensates. *Dev. Cell.* **55**(1), 30–44.
11. Jiang H., Wang S., Huang Y., He X., Cui H., Zhu X., Zheng Y. (2015) Phase transition of spindle-associated protein regulate spindle apparatus assembly. *Cell.* **163**(1), 108–122.
12. Huang Y., Li T., Ems-McClung S.C., Walczak C.E., Prigent C., Zhu X., Zhang X., Zheng Y. (2018) Aurora A activation in mitosis promoted by BuGZ. *J. Cell. Biol.* **217**(1), 107–116.
13. Lee K.H., Zhang P., Kim H.J., Mitrea D.M., Sarkar M., Freibaum B.D., Cika J., Coughlin M., Messing J., Molliex A., Maxwell B.A., Kim N.C., Temirov J., Moore J., Kolaitis R.M., Shaw T.I., Bai B., Peng J., Kriwacki R.W., Taylor J.P. (2016) C9orf72 dipeptide repeats impair the assembly, dynamics, and function of membrane-less organelles. *Cell.* **167**(3), 774–788. e717.
14. Altmeyer M., Neelsen K.J., Teloni F., Pozdnyakova I., Pellegrino S., Grøfte M., Rask M.D., Streicher W., Jungmichel S., Nielsen M.L., Lukas J. (2015) Liquid demixing of intrinsically disordered proteins is seeded by poly(ADP-ribose). *Nat. Commun.* **6**, 8088.
15. Boeynaems S., Alberti S., Fawzi N.L., Mittag T., Polyimenidou M., Rousseau F., Schymkowitz J., Shorter J., Wolozin B., Van Den Bosch L., Tompa P., Fuxreiter M. (2018) Protein phase separation: a new phase in cell biology. *Trends Cell Biol.* **28**(6), 420–435.
16. Shin Y., Brangwynne C.P. (2017) Liquid phase condensation in cell physiology and disease. *Science.* **357**, 6357.
17. Patel A., Lee H.O., Jawerth L., Maharana S., Jahnke M., Hein M.Y., Stoyanov S., Mahamid J., Saha S., Franzmann T.M., Pozniakowski A., Poser I., Maghelli N., Royer L.A., Weigert M., Myers E.W., Grill S., Drechsel D., Hyman A.A., Alberti S. (2015) A liquid-to-solid phase transition of the ALS protein FUS accelerated by disease mutation. *Cell.* **162**(5), 1066–1077.
18. Pannone B.K., Kim S.D., Noe D.A., Wolin S.L. (2001) Multiple functional interactions between components of the Lsm2-Lsm8 complex, U6 snRNA, and the yeast La protein. *Genetics.* **158**(1), 187–196.
19. Arribas-Layton M., Dennis J., Bennett E.J., Damgaard C.K., Lykke-Andersen J. (2016) The C-terminal RGG domain of human Lsm4 promotes processing body formation stimulated by arginine dimethylation. *Mol. Cell. Biol.* **36**(17), 2226–2235.
20. Reijns M.A., Alexander R.D., Spiller M.P., Beggs J.D. (2008) A role for Q/N-rich aggregation-prone regions in P-body localization. *J. Cell Sci.* **121**(Pt 15), 2463–2472.
21. Roth A.J., Shuman S., Schwer B. (2018) Defining essential elements and genetic interactions of the yeast Lsm2-8 ring and demonstration that essentiality of Lsm2-8 is bypassed via overexpression of U6 snRNA or the U6 snRNP subunit Prp24. *RNA.* **24**(6), 853–864.
22. Lyons S.M., Ricciardi A.S., Guo A.Y., Kambach C., Marzluff W.F. (2014) The C-terminal extension of Lsm4 interacts directly with the 3' end of the histone mRNP and is required for efficient histone mRNA degradation. *RNA.* **20**(1), 88–102.
23. Decker C.J., Teixeira D., Parker R. (2007) Edc3p and a glutamine/asparagine-rich domain of Lsm4p function in processing body assembly in *Saccharomyces cerevisiae*. *J. Cell Biol.* **179**(3), 437–449.
24. Mennie A.K., Moser B.A., Nakamura T.M. (2018) LARP7-like protein Pof8 regulates telomerase assembly and poly(A)+ TERRA expression in fission yeast. *Nat. Commun.* **9**(1), 1–12.
25. Adamson B.S., Smogorzewska A., Sigoillot F.D., King R.W., Elledge S.J. (2012) A genome-wide study of homologous recombination in mammalian cells identifies RBMX, a novel component of the DNA damage response. *Nat. Cell Biol.* **14**(3), 318–328.
26. Alabrudzinska M., Skoneczny M., Skoneczna A. (2011) Diploid-specific genome stability genes of *S. cerevisiae*: genomic screen reveals haploidization as an escape from persisting DNA rearrangement stress. *PLoS One.* **6**(6), e21124.
27. Lin Y., Protter D.S., Rosen M.K., Parker R. (2015) Formation and maturation of phase-separated liquid droplets by RNA-binding proteins. *Mol. Cell.* **60**(2), 208–219.
28. Linding R., Jensen L.J., Diella F., Bork P., Gibson T.J., Russell R.B. (2003) Protein disorder prediction: implications for structural proteomics. *Structure.* **11**(11), 1453–1459.
29. Schuster B.S., Reed E.H., Parthasarathy R., Jahnke C.N., Caldwell R.M., Bermudez J.G., Ramage H., Good M.C., Hammer D.A. (2018) Controllable protein phase separation and modular recruitment to form responsive membraneless organelles. *Nat. Commun.* **9**(1), 2985.
30. Uversky V.N. (2017) Intrinsically disordered proteins in overcrowded milieu: membrane-less organelles, phase separation, and intrinsic disorder. *Curr. Opin. Struct. Biol.* **44**, 18–30.
31. Sasahara K., McPhie P., Minton A.P. (2003) Effect of dextran on protein stability and conformation attributed to macromolecular crowding. *J. Mol. Biol.* **326**(4), 1227–1237.
32. Alberti S., Saha S., Woodruff J.B., Franzmann T.M., Wang J., Hyman A.A. (2018) A user's guide for phase

- separation assays with purified proteins. *J. Mol. Biol.* **430**(23), 4806–4820.
33. Alberti S., Gladfelter A., Mittag T. (2019) Considerations and challenges in studying liquid–liquid phase separation and biomolecular condensates. *Cell.* **176**(3), 419–434.
 34. Verdone L., Galardi S., Page D., Beggs J.D. (2004) Lsm proteins promote regeneration of pre-mRNA splicing activity. *Curr. Biol.* **14**(16), 1487–1491.
 35. Rao B.S., Parker R. (2017) Numerous interactions act redundantly to assemble a tunable size of P bodies in *Saccharomyces cerevisiae*. *Proc. Natl. Acad. Sci. USA.* **114**(45), E9569–E9578.
 36. Tang W., Kannan R., Blanchette M., Baumann P. (2012) Telomerase RNA biogenesis involves sequential binding by Sm and Lsm complexes. *Nature.* **484**(7393), 260–264.

Phase Separation of Purified Human LSM4 Protein

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Liquid–liquid phase separation of proteins occur in a number of biological processes, such as regulation of transcription, processing, and RNA maturation. Sm-like protein 4 (LSM4) is involved in multiple processes, including pre-mRNA splicing and P-bodies assembly. Before investigating the involvement of LSM4 in the separation of the two liquid phases during RNA processing or maturation, the separation of the liquid phases in an *in vitro* preparation of LSM4 protein should be first be detected. The mCherry-LSM4 plasmid was derived from pET30a and used to isolate mCherry-LSM4 protein from prokaryotic cells (*Escherichia coli* strain BL21). The mCherry-LSM4 protein was purified using Ni-NTA resin. The protein was further purified by fast protein liquid chromatography. Delta-Vision wide-field fluorescence microscopy was used to observe the dynamic liquid–liquid phase separation of the LSM4 protein *in vitro*. Analysis of the LSM4 protein structure using the Predictor of Natural Disordered Regions database revealed that its C-terminus contains a low complexity domain. A purified preparation of full-length human LSM4 protein was obtained from *E. coli*. Human LSM4 was shown to provide concentration-dependent separation of liquid–liquid phases *in vitro* in buffer with crowding reagents. Salts in high concentration and 1,6-hexanediol block the LSM4-induced separation of the two liquid phases. In addition, *in vitro* fusion of LSM4 protein droplets is observed. These results indicate that the full-length human LSM4 protein has the ability to form liquid inclusions and induce liquid–liquid phase separation *in vitro*.

Keywords: liquid–liquid phase separation, Sm-like protein 4, fast protein liquid chromatography, protein purification, biophysical process