

ИДЕНТИФИКАЦИЯ ФРАГМЕНТА СТИГМАСПЕЦИФИЧНОЙ ЭКСПРЕССИИ В ПРОМОТОРЕ ГЕНА ХИТИНАЗЫ КЛАССА I СОИ¹

© 2023 г. С. М. Zhao^a, Н. Hou^a, М. G. Xing^b, R.-G. Xue^{a, *}

^aCollege of Life Sciences, Qingdao Agricultural University, Qingdao, 266109 China

^bUbrigene (Jinan) Biosciences Co., Ltd, Jinan, 250000 China

*e-mail: xuerengao@163.com

Поступила в редакцию 26.04.2022 г.

После доработки 01.08.2022 г.

Принята к публикации 23.08.2022 г.

Уровень экспрессии гетерологичных генов в трансгенных растениях служит важным показателем эффективности работы генов. Тонкая настройка экспрессии трансгенов ограничена небольшим реperтуаром известных на данный момент эффективных промоторов. Нами клонирован и охарактеризован тканеспецифичный фрагмент промотора гена хитиназы класса I сои (*GmChi1*). Промотор *GmChi1*(*GmChi1P*) клонировали из сои сорта *Jungery*. Последовательность промотора содержит ряд предполагаемых *цис*-действующих элементов, включая тканеспецифичные мотивы и мотивы, регулируемые в условиях стресса. По результатам гистохимического анализа самая высокая активность репортерного фермента β -глюкуронидазы (GUS), находящегося под контролем *GmChi1P*, обнаружена в корнях трансгенного растения, *Nicotiana tabacum* cv. NC89, на стадии формирования ростка с четырьмя листьями. Интересно, что обработка салициловой кислотой эффективно подавляла высокую активность GUS в корнях трансгенного табака. Делеционным анализом *GmChi1P* установлено, что последовательности, расположенные между позициями -719 и -382, содержат ключевые *цис*-элементы, ответственные за экспрессию репортерного гена *uidA* (кодирующего GUS) в листьях, корнях и местах поранения *Nicotiana tabacum*. Кроме того, согласно результатам флуорометрического анализа, активность укороченных промоторов: от *ChiP*(-1292) до *ChiP*(-719) – в корнях трансгенного табака значительно подавляется абсцизовой кислотой и полностью – салициловой. Обнаружено также, что промотор *ChiP*(-382) экспрессируется исключительно в рильце цветков трансгенного растения. С использованием репортерного фермента GUS не обнаружено окрашивания в других органах цветка трансгенного *Nicotiana tabacum*, включая чашелистики, лепестки, пыльники, нити и завязи, а также ни в одной из вегетативных тканей. Полученные результаты свидетельствуют о том, что фрагмент промотора *ChiP*(-382) может быть использован в тканеспецифичной регуляции экспрессии генов и генной инженерии растений.

Ключевые слова: *GmChi1*, соя, промотор, трансгенные растения, *Nicotiana tabacum*, стигмаспецифичный промотор

DOI: 10.31857/S0026898423010172, **EDN:** AXAYSA

СПИСОК ЛИТЕРАТУРЫ

- Li W., Yu D., Yu J., Zhu D., Zhao Q. (2018) Functional analysis of maize silk-specific *ZmbZIP25* promoter. *Int. J. Mol. Sci.* **19**, 822–835.
- Bowles D.J. (1990) Defense related proteins in higher plants. *Annu. Rev. Biochem.* **59**, 873–907.
- Kenton P., Darby R.M., Shelley G., Draper J. (2000) A *PR5* gene promoter from *Asparagus officinalis* (AoPRT-L) is not induced by abiotic stress, but is activated around sites of pathogen challenge and salicylate in transgenic tobacco. *Mol. Plant Pathol.* **1**, 367–378.
- Brown R.L., Kazan K., McGrath K.C., Maclean D.J., Manners J.M. (2003) A role for the GCC-box in jasmonate-mediated activation of the *PDF1.2* gene of *Arabidopsis*. *Plant Physiol.* **132**, 1020–1032.
- Collinge D.B., Kragh K.M., Mikkelsen J.D., Rasmussen U., Vad K. (1993) Plant chitinases. *Plant J.* **3**, 31–40.
- Sticken M.B., Graham L.S. (1994) Plant chitinases. *Can. J. Bot.* **72**, 1057–1083.
- Cletus J., Balasubramanian V., Vashisht D., Sakthivel N. (2013) Transgenic expression of plant chitinases to en-

¹ Статья представлена авторами на английском языке.

- hance disease resistance. *Biotechnol. Lett.* **35**, 1719–1732.
8. Abd El-Rahman S.S., Mazen M.M., Mohamed H.I., Mahmoud N.M. (2012) Induction of defence related enzymes and phenolic compounds in lupin (*Lupinus albus* L.) and their effects on host resistance against *Fusarium* wilt. *Eur. J. Plant Pathol.* **134**, 105–116.
 9. Acharya K., Chakraborty N., Dutta A.K., Sarkar S., Acharya R. (2011) Signaling role of nitric oxide in the induction of plant defense by exogenous application of abiotic inducers. *Arch. Phytopath. Plant Protect.* **44**, 1501–1511.
 10. Arlorio M., Ludwig A., Boller T., Bonfante P. (1992) Inhibition of fungal growth by plant chitinases and β -1,3-glucanases. *Protoplasma* **171**, 34–43.
 11. Veluthakkal R., Dasgupta M.G. (2012) Isolation and characterization of pathogen defence-related class I chitinase from the actinorrhizal tree *Casuarina equisetifolia*. *For. Path.* **42**, 467–480.
 12. Yeboah N.A., Arahira M., Nong V.H., Zhang D., Kadokura K., Watanabe A., Fukazawa C. (1998) A class III acidic endochitinase is specifically expressed in the developing seeds of soybean (*Glycine max* [L.] Merr.). *Plant Mol. Biol.* **36**, 407–415.
 13. Gijzen M., Kuflu K., Qutob D., Chernys J.T. (2001) A class I chitinase from soybean seed coat. *J. Exp. Bot.* **52**, 2283–2289.
 14. Zhou G. (2017) Cloning and functional analysis of specific promoter p51408 in stigma exertion of wild rice (*Oryza rufipogon* Griff.) from yuanjiang city. *Southwest China J. Agricult. Sci.* **11**, 2393–2397.
 15. Yu L., Ma T., Zhang Y., Hu Y., Yu K., Chen Y., Ma H., Zhao J. (2017) Identification and analysis of the stigma and embryo sac-preferential/specific genes in rice pistils. *BMC Plant Biology* **17**, 60–81.
 16. Stein J.C., Dixit R., Nasrallah M.E., Nasrallah J.B. (1996) SRK, the stigma-specific *Slocus* receptor kinase of *Brassica*, is targeted to the plasma membrane in transgenic tobacco. *Plant Cell* **8**, 429–445.
 17. Hackett R.M., Cadwallader G., Franklin F.C. (1996) Functional analysis of a *Brassica oleracea* SLR1 gene promoter. *Plant Physiol.* **112**, 1601–1607.
 18. Alvim F.C., Carolino S.M., Cascardo J.C., Nunes C.C., Martinez C.A., Otoni W.C., Fontes E.P. (2001) Enhanced accumulation of BiP in transgenic plants confers tolerance to water stress. *Plant Physiol.* **126**, 1042–1054.
 19. Higo K., Ugawa Y., Iwamoto M., Korenaga T. (1999) Plant *cis*-acting regulatory DNA element (PLACE) database. *Nucleic Acids Res.* **27**, 297–300.
 20. Jefferson R.A., Kavanagh T.A., Bevan M.W. (1987) GUS fusions: β -glucuronidase as a sensitive and versatile gene fusion marker in higher plants. *EMBO J.* **6**, 3901–3907.
 21. Xu X., Chen C., Fan B., Chen Z. (2006) Physical and functional interactions between pathogen-induced *Arabidopsis* WRKY18, WRKY40, and WRKY60 transcription factors. *Plant Cell* **18**, 1310–1326.
 22. Nishiuchi T., Shinshi H., Suzuki K. (2004) Rapid and transient activation of transcription of the *ERF3* gene by wounding in tobacco leaves: possible involvement of NtWRKYs and autorepression. *J. Biol. Chem.* **279**, 55355–55361.
 23. Chen C., Chen Z. (2002) Potentiation of developmentally regulated plant defense response by AtWRKY18, a pathogen-induced *Arabidopsis* transcription factor. *Plant Physiol.* **129**, 706–716.
 24. Zhang Z.L., Xie Z., Zou X., Casaretto J., Ho T.H., Shen Q.J. (2004) A rice WRKY gene encodes a transcriptional repressor of the gibberellin signaling pathway in aleurone cells. *Plant Physiol.* **134**, 1500–1513.
 25. Xie Z., Zhang Z.L., Zou X., Huang J., Ruas P., Thompson D., Shen Q.J. (2005) Annotations and functional analyses of the rice WRKY gene superfamily reveal positive and negative regulators of abscisic acid signaling in aleurone cells. *Plant Physiol.* **137**, 176–189.
 26. Urao T., Yamaguchi-Shinozaki K., Urao S., Shinozaki K. (1993) An *Arabidopsis myb* homolog is induced by dehydration stress and its gene product binds to the conserved MYB recognition sequence. *Plant Cell* **5**, 1529–1539.
 27. Solano R., Nieto C., Avila J., Canas L., Diaz I., Paz-Ares J. (1995) DNA binding specificity of a petal epidermis-specific MYB transcription factor (MYB.Ph3) from *Petunia* hybrid. *EMBO J.* **14**, 1773–1784.
 28. Agarwal M., Hao Y., Kapoor A., Dong C.H., Fujii H., Zheng X., Zhu J.K. (2006) A R2R3 type MYB transcription factor is involved in the cold regulation of CBF genes and in acquired freezing tolerance. *J. Biol. Chem.* **281**, 37636–37645.
 29. Hartmann U., Sagasser M., Mehrtens F., Stracke R., Weißhaar B. (2005) Differential combinatorial interactions of cis-acting elements recognized by R2R3-MYB, BZIP, and BHLH factors control light-responsive and tissue-specific activation of phenylpropanoid biosynthesis genes. *Plant Mol. Biol.* **57**, 155–171.
 30. Park H.C., Kim M.L., Kang Y.H., Jeon J.M., Yoo J.H., Kim M.C., Park C.Y., Jeong J.C., Moon B.C., Lee J.H., Yoon H.W., Lee S.H., Chung W.S., Lim C.O., Lee S.Y., Hong J.C., Cho M.J. (2004) Pathogen- and NaCl-induced expression of the SCaM-4 promoter is mediated in part by a GT-1 box that interacts with a GT-1-like transcription factor. *Plant Physiol.* **135**, 2150–2161.
 31. Luo H., Song F., Goodman R.M., Zheng Z. (2005) Up-regulation of *OsBIHD1*, a rice gene encoding BELL homeodomain transcriptional factor, in disease resistance responses. *Plant Biol. (Stuttg.)* **7**, 459–468.
 32. Nakashima K., Fujita Y., Katsura K., Maruyama K., Narusaka Y., Seki M., Shinozaki K., Yamaguchi-Shinozaki K. (2006) Transcriptional regulation of ABI3- and ABA-responsive genes including *RD29B* and *RD29A* in seeds, germinating embryos, and seedlings of *Arabidopsis*. *Plant Mol. Biol.* **60**, 51–68.
 33. Simpson S.D., Nakashima K., Narusaka Y., Seki M., Shinozaki K., Yamaguchi-Shinozaki K. (2003) Two different novel *cis*-acting elements of *erd1*, a *clpA* homologous *Arabidopsis* gene function in induction by dehydration stress and dark-induced senescence. *Plant J.* **33**, 259–270.
 34. Fehlberg V., Vieweg M.F., Dohmann E.M., Hohnjec N., Puhler A., Perlick A.M., Kuster H. (2005) The promoter of the leghaemoglobin gene *VfLb29*: functional analysis and identification of modules necessary for its acti-

- vation in the infected cells of root nodules and in the arbuscule-containing cells of mycorrhizal roots. *J. Exp. Bot.* **56**, 799–806
35. Vieweg M.F., Fröhling M., Quandt H.J., Heim U., Bäumlein H., Pühler A., Küster H., Perlitz A.M. (2004) The promoter of the *Vicia faba* L. leghemoglobin gene *VfLb29* is specifically activated in the infected cells of root nodules and in the arbuscule-containing cells of mycorrhizal roots from different legume and nonlegume plants. *MPMI*. **17**, 62–69.
 36. Kagaya Y., Ohmiya K., Hattori T. (1999) RAV1, a novel DNA-binding protein, binds to bipartite recognition sequence through two distinct DNA-binding domains uniquely found in higher plants. *Nucleic Acids Res.* **27**, 470–478.
 37. Gowik U., Burscheidt J., Akyildiz M., Schlue U., Koczor M., Streubel M., Westhoff P. (2004) *cis*-Regulatory elements for mesophyll-specific gene expression in the C₄ plant *Flaveria trinervia*, the promoter of the C₄ phosphoenolpyruvate carboxylase gene. *Plant Cell*. **16**, 1077–1090.
 38. Filichkin S.A., Leonard J.M., Monteros A., Liu P.P., Nonogaki H. (2004) A novel endo-β-mannanase gene in tomato LeMAN5 is associated with anther and pollen development. *Plant Physiol.* **134**, 1080–1087.
 39. Yanagisawa S. (2000) Dof1 and Dof2 transcription factors are associated with expression of multiple genes involved in carbon metabolism in maize. *Plant J.* **21**, 281–288.
 40. Niki T., Mitsuhashi I., Seo S., Ohtsubo N., Ohashi Y. (1998) Antagonistic effect of salicylic acid and jasmonic acid on the expression of pathogenesis-related (PR) protein genes in wounded mature tobacco leaves. *Plant Cell Physiol.* **39**, 500–507.
 41. Chen L., Jiang B., Wu C., Sun S., Hou W., Han T. (2014) *GmPRP2* promoter drives root-preferential expression in transgenic *Arabidopsis* and soybean hairy roots. *BMC Plant Biol.* **14**, 245–257.
 42. Lalonde B.A., Nasrallah M.E., Dwyer K.G., Chen C.H., Barlow B., Nasrallah J. (1989) A highly conserved *Brassica* gene with homology to the S-locus specific glycoprotein structural gene. *Plant Cell*. **1**, 249–258.
 43. Ke L., Zheng T., Wu X., Chen J., Zhu S. (2007) Isolation and sequence analysis of the stigma-specific promoter from *Brassica napus*. *J. Agricult. Biotechnol.* **15**, 257–262.

Identification of Stigma-Specific Expression Fragment in the Promoter of the Soybean Chitinase Class I Gene

C. M. Zhao¹, H. Hou¹, M. G. Xing², and R.-G. Xue^{1,*}

¹College of Life Sciences, Qingdao Agricultural University, Qingdao, 266109 China

²Ubrigene (Jinan) Biosciences Co., Ltd, Jinan, 250000 China

*e-mail: xuerengao@163.com

The expression level of heterologous genes in transgenic plants serves as an important indicator of gene efficiency. The small number of currently known effective promoters, limits the possibilities in fine-tuning the expression of transgenes. We cloned and characterized a tissue-specific promoter fragment of the soybean chitinase class I gene (*GmChi1*). The *GmChi1* promoter (*GmChi1P*) was cloned from Jungery soybean. The promoter sequence contains a number of putative *cis*-acting elements, including tissue-specific and stress-regulated motifs. By histochemical analysis, the *GmChi1P*-controlled β-glucuronidase (GUS) reporter enzyme activity was shown to be highest in the roots of transgenic *Nicotiana tabacum* cv. NC89 at the four-leaf sprout formation stage. Interestingly, the high GUS activity in transgenic tobacco roots was effectively suppressed by salicylic acid (SA) treatment. Deletion analysis of *GmChi1P* revealed that the sequences located between positions –719 and –382 contain key *cis*-elements responsible for the reporter *uidA* gene expression (encoding GUS) in leaves, roots, and wounds of *Nicotiana tabacum*. In addition, fluorometric analysis showed that the activity of the shortened *ChiP*(–1292) to *ChiP*(–719) promoters in the roots of transgenic tobacco was significantly suppressed by abscisic acid and completely suppressed by SA. The *ChiP*(–382) promoter was also found to be expressed exclusively in the stigma of transgenic tobacco flowers. Using the GUS reporter enzyme, no staining was detected in other flower organs in transgenic *Nicotiana tabacum*, including sepals, petals, anthers, filaments, and ovaries, or in any vegetative tissues. The results indicate that the promoter fragment *ChiP*(–382) can be used in tissue-specific regulation of gene expression and plant genetic engineering.

Keywords: *GmChi1*, soybean, promoter, transgenic plant, *Nicotiana tabacum*, stigma-specific promoter